

Fully Automated QuEChERS for Organochlorine and Organophosphate Pesticides in Tomato Juice and Red Wine

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PAL SYSTEM
Ingenious sample handling

Overview

Using PAL Robotic Tool Change (RTC) system, a fully automated QuEChERS was developed for the extraction and clean-up of organochlorine and organophosphate pesticides from homogeneous samples. The automated QuEChERS workflow includes extraction with acetonitrile, salting out with saturated sodium chloride solution and the clean-up with PAL μ SPE prior to injection into the GC-MS/MS for analysis. Method validations were achieved by using the automated matrix matches calibration spiked from 1 ng/mL to 100 ng/mL of organochlorine and organophosphate pesticide standards into the PAL μ SPE-cleaned tomato juice and red wine samples. The matrix match calibration curve linearities were at R^2 0.995 or better. By spiking 10 ng/mL of pesticide standards into a raw tomato juice and red wine, the recoveries were obtained in the range of 70% - 130%, with $n = 6$ samples to determine the automated workflow precisions.

Introduction

QuEChERS is the quick, easy, cheap effective, rugged and safe sample preparation method developed by M. Anastassiades and S.J. Lehotay in 2003. Since then, this technique has become the widely used sample preparation approach in pesticide residue analyses. According to the QuEChERS website, about 45 min are needed to manually prepare 8 samples in the laboratory [1]. In the QuEChERS method, acetonitrile is used as extraction solvent, followed by adding NaCl and buffer salts. After shaking and centrifugation traditionally dispersive solid phase extraction (dSPE) is used for extract clean-up before analysis by GC-MS or LC-MS. High matrix load in GC-MS, the matrix effects in LC-MS, enhancing or suppressing analyte signals, varying extraction recoveries were major challenges that were approached to overcome by many matrix specific modified clean-up agents and procedures.

Analytical Strategy

Using PAL μ SPE in Matrix Clean-Up

A cartridge based miniaturized solid phase extraction, known as PAL μ SPE, was fully automated by using the PAL RTC system to perform the fully automated QuEChERS preparation workflow.

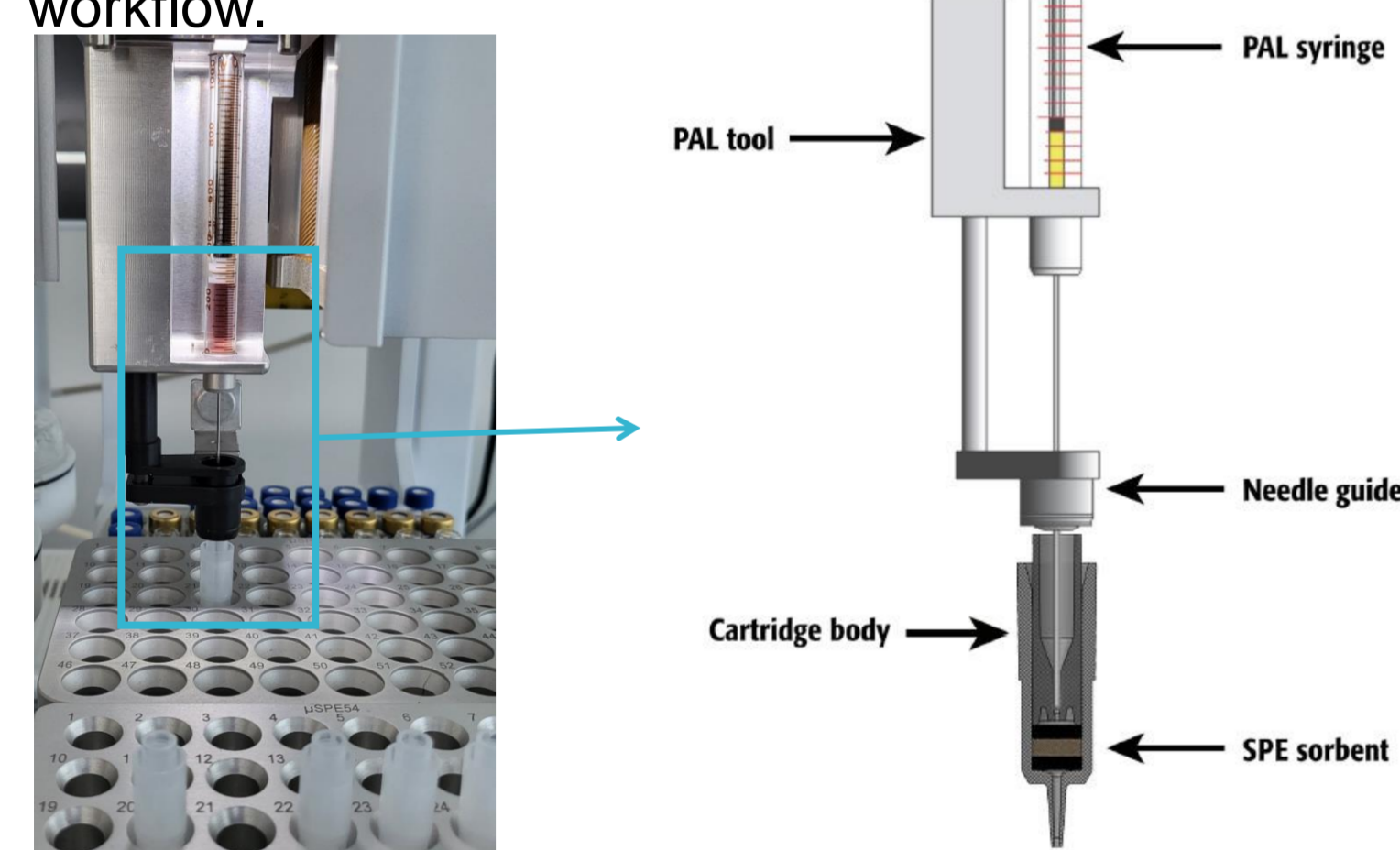


Figure 1. Principle of the PAL μ SPE clean-up

An optimized sorbent bed for GC QuEChERS analysis, containing a mixture of 45 mg of $MgSO_4$, PSA, C18EC and CarbonX was filled by CTC Analytics into proprietary PAL μ SPE cartridges. This PAL μ SPE cartridges work in scavenging mode to retain the sample matrix inside the sorbent bed. A clean extract delivering the pesticide compounds is eluted into empty vials beneath the cartridges for online injection to GC-MS.



Figure 2. Red wine sample, before and after the PAL μ SPE clean-up

Instrument Setup

The fully automated workflow, including sample blank, post spike with calibration and internal standards was carried out with the instrument setup as shown in Figure 3. The automated QuEChERS was established by the PAL RTC system with a vortexing module, solvent module to store up to 100 mL of acetonitrile and saturated sodium chloride solution, a fast wash module with acetonitrile and water for active syringe wash. In the Tray Holder, as shown in Figure 3, rack 1 was used to place the tomato juice and red wine samples in 2 mL vials. The clean up cartridges were placed on the dedicated cartridge holder in rack 3. Empty vials with slitted septa were placed at the center rack 2 underneath of the aluminium vial cover to receive cleaned extract.

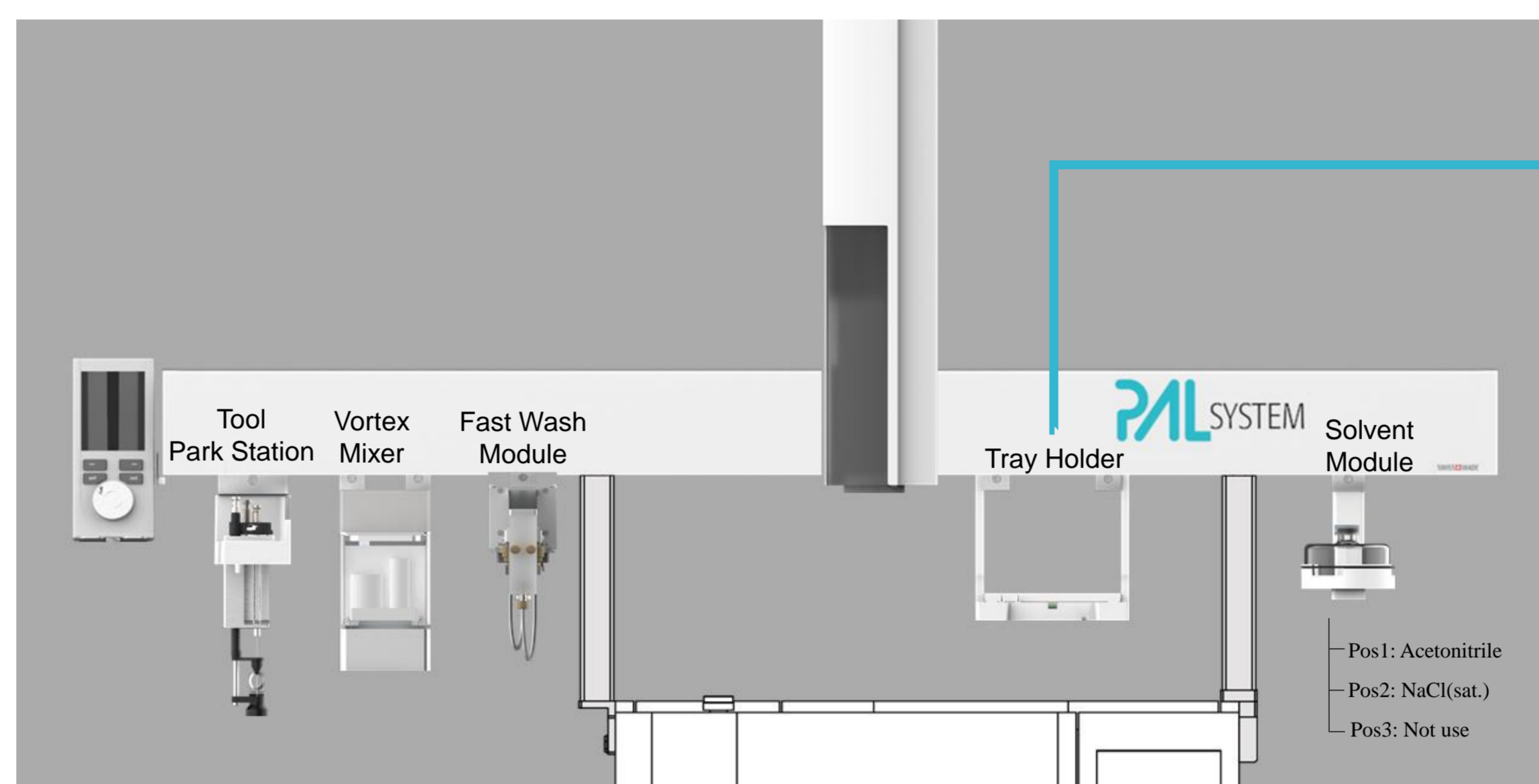
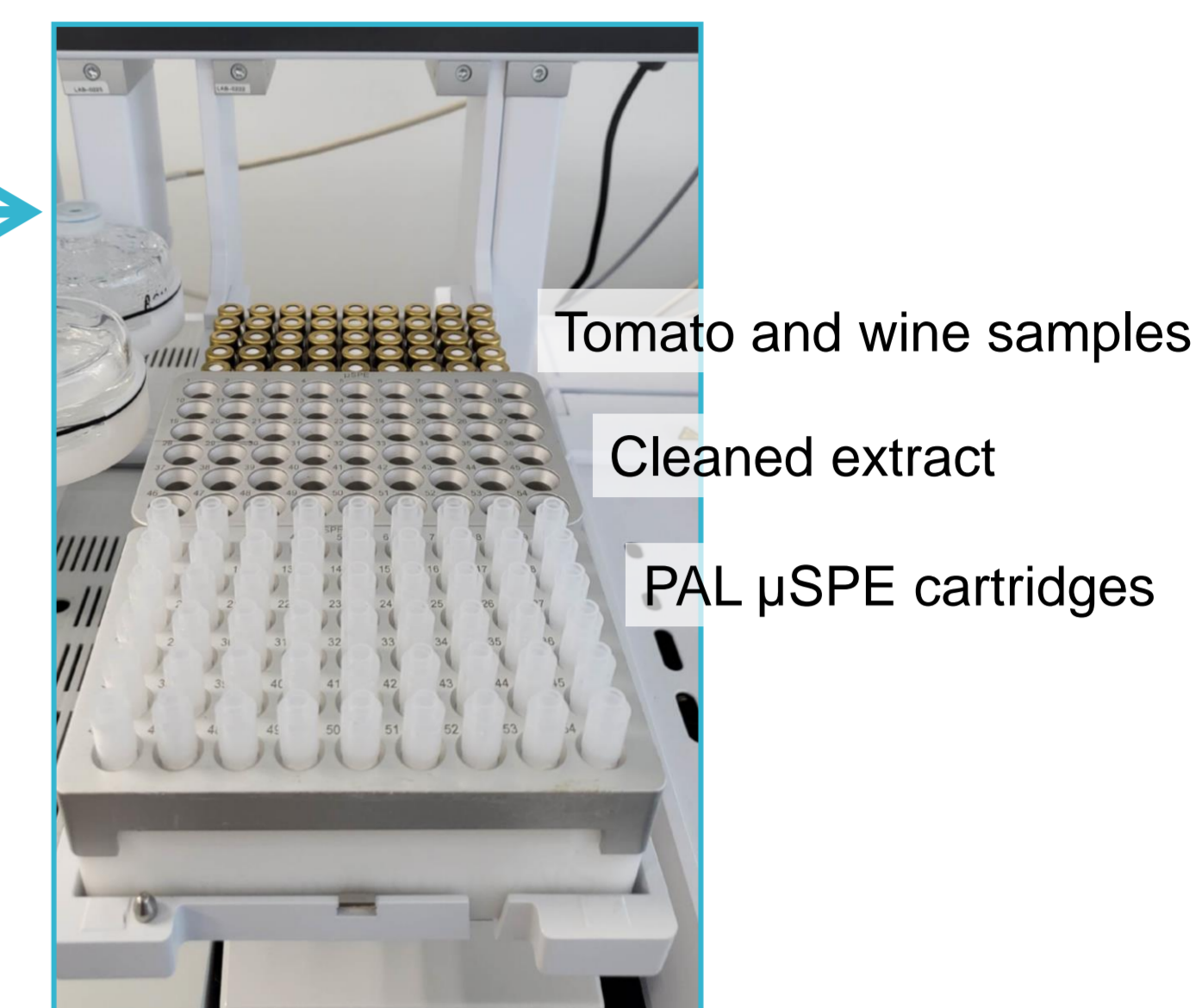


Figure 3. PAL RTC system configuration for the fully automated QuEChERS analysis



Automated QuEChERS Workflow in Analyzing Tomato Juice and Wine Samples

The only manual step in the automated QuEChERS workflow was a pipetting of 500 μ L of sample into 2 mL autosampler vials. The subsequent steps such as adding Acetonitrile, salt solution and clean-up by PAL μ SPE were carried out automatically by the PAL RTC system based on the workflow shown in Figure 4. The organic and aqueous phases were well separated after adding in the saturated salt solution. No centrifugation was required. The PAL μ SPE cartridge with $MgSO_4$, PSA, C18EC and CarbonX sorbents provided a high clean-up efficiency, as shown in Figure 5 for tomato juice and Figure 6 for red wine samples.

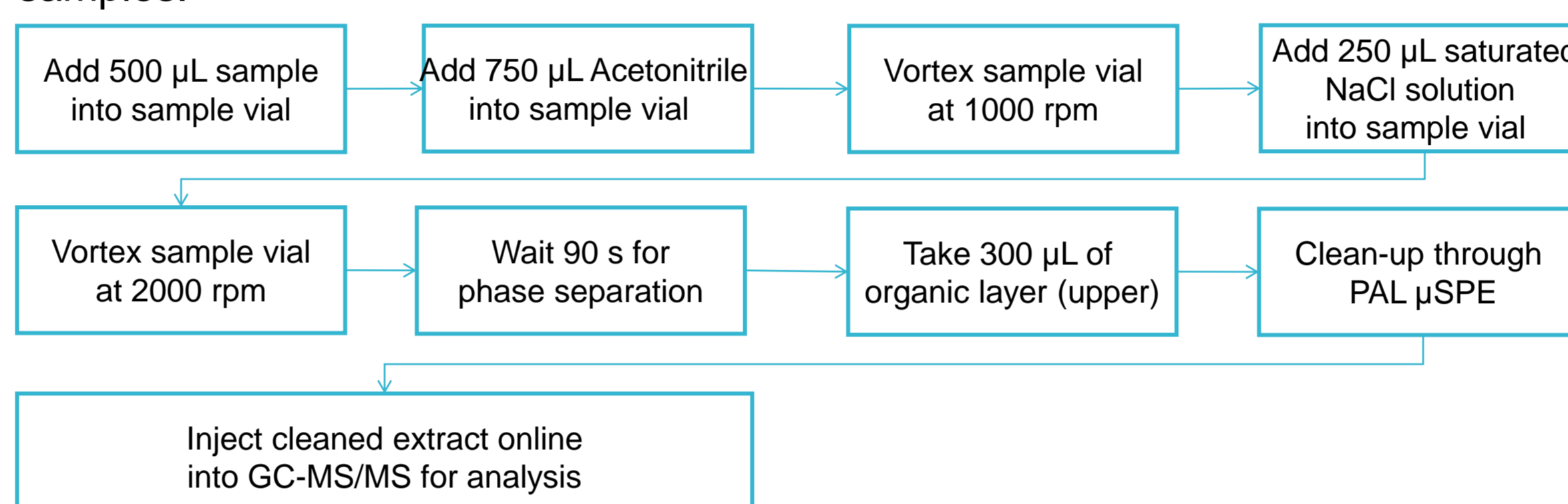


Figure 4. Automated QuEChERS workflow by PAL RTC System

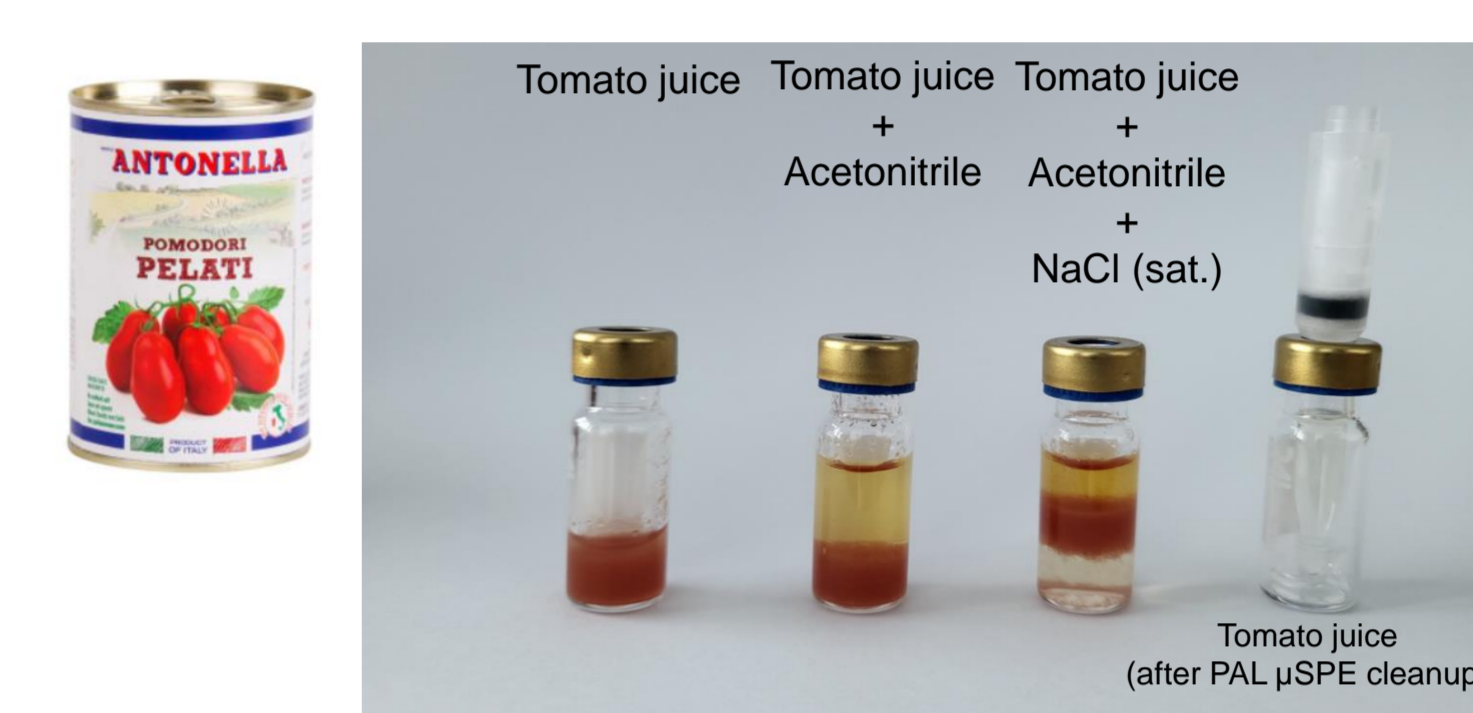


Figure 5. QuEChERS in tomato juice

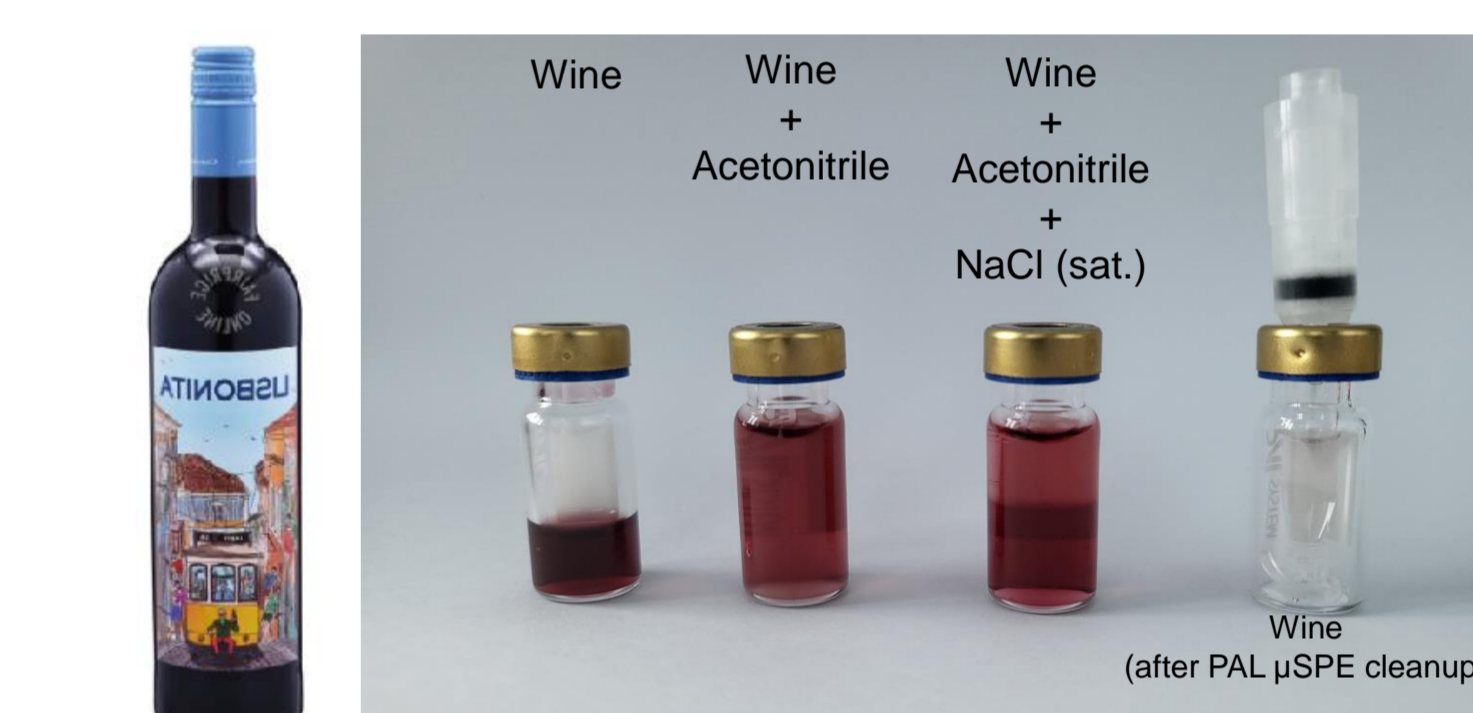


Figure 6. QuEChERS in wine

Results

Prior to analyzing the tomato juice and red wine samples, a Shimadzu GCMS-TQ8040, equipped with Rxi-5Sil MS (30 m x 0.25 mm x 0.25 μ m) capillary column was optimized to achieve the detection limit to at least 1 ng/mL. A full MRM total ion chromatogram (TIC) of the 100 ng/mL organochlorine and organophosphate standards is shown as Figure 7. Upon completed QuEChERS extraction and clean-up, the tomato juice and wine samples were injected at 3 μ L into this optimized GC-MS/MS system for analysis.

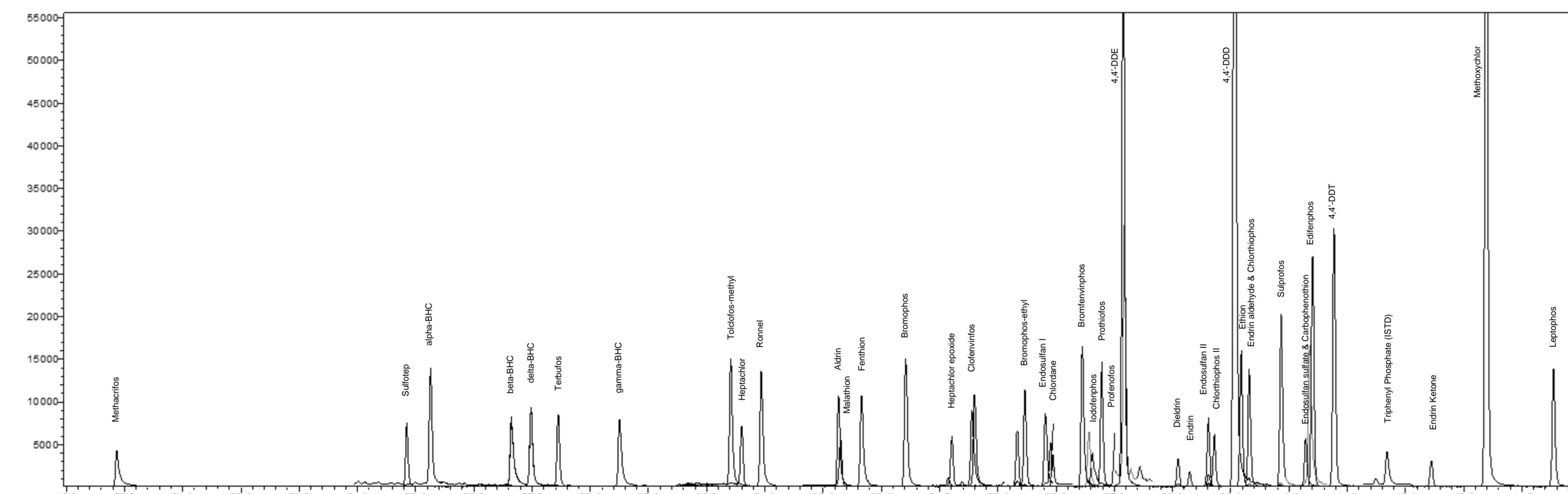


Figure 7. Total Ion Chromatograms (TICs) of 100ng/mL organochlorine and organophosphate pesticide standards by the GC-MS/MS.

| Compounds (based on elution order) | Tomato Juice | | | Red Wine | | |
|--|--------------|-------------------------|----------|-----------|-------------------------|----------|
| | Linearity | Precision %RSD (n=5) | Recovery | Linearity | Precision %RSD (n=6) | Recovery |
| Methacrifos | 0.9995 | 8% | 129% | 0.9984 | 24% | 103% |
| Sulfotep | 0.9990 | 7% | 94% | 0.9982 | 16% | 87% |
| alpha-BHC | 0.9977 | 13% | 104% | 0.9924 | 20% | 101% |
| beta-BHC | 0.9956 | 6% | 115% | 0.9958 | 12% | 115% |
| delta-BHC | 0.9966 | 7% | 118% | 0.9956 | 11% | 115% |
| Terbufos | 0.9994 | 8% | 108% | 0.9953 | 14% | 81% |
| gamma-BHC | 0.9991 | 9% | 87% | 0.9979 | 13% | 113% |
| Toxicofos-methyl | 0.9998 | 5% | 107% | 0.9998 | 10% | 114% |
| Heptachlor | 0.9972 | 5% | 99% | 0.9969 | 20% | 84% |
| Ronnel | 0.9990 | 4% | 105% | 0.9970 | 10% | 112% |
| Aldrin | 0.9989 | 15% | 85% | 0.9973 | 21% | 72% |
| Malathion | 0.9972 | 9% | 86% | 0.9960 | 13% | 115% |
| Fenthion | 0.9964 | 10% | 112% | 0.9986 | 11% | 106% |
| Bromophos | 0.9990 | 8% | 104% | 0.9994 | 9% | 109% |
| Heptachlor epoxide | 0.9961 | 13% | 84% | 0.9973 | 13% | 86% |
| Clofeninfos | 0.9608 | 12% | 89% | 0.9971 | 13% | 111% |
| Bromophos-ethyl | 0.9977 | 3% | 86% | 0.9995 | 8% | 68% |
| Endosulfan I | 0.9959 | 5% | 79% | 0.9997 | 16% | 77% |
| Chlordane | 0.9970 | 4% | 115% | 0.9985 | 17% | 81% |
| Bromofeniphos | 0.9761 | 13% | 80% | 0.9989 | 28% | 103% |
| Iodofeniphos | 0.9966 | 18% | 91% | 0.9993 | 16% | 82% |
| Prothiofos | 0.9989 | 10% | 79% | 0.9998 | 14% | 69% |
| 4,4'-DDE | 0.9982 | 15% | 83% | 0.9997 | 19% | 86% |
| Profenofos | 0.9865 | 10% | 86% | 0.9995 | 11% | 94% |
| Dieldrin | 0.9965 | 6% | 112% | 0.9985 | 18% | 75% |
| Endrin | 0.9968 | 12% | 84% | 0.9993 | 14% | 101% |
| Endosulfan II | 0.9984 | 6% | 94% | 0.9981 | 14% | 107% |
| Chlorthiophos III | 0.9982 | 11% | 76% | 0.9988 | 9% | 64% |
| 4,4'-DDD | 0.9988 | 6% | 102% | 0.9985 | 15% | 88% |
| Ethion | 0.9980 | 6% | 111% | 0.9993 | 14% | 83% |
| Endrin aldehyde | 0.9988 | 3% | 55% | 0.9712 | 30% | 91% |
| Chlorthiophos | 0.9956 | 11% | 81% | 0.9992 | 13% | 61% |
| Sulprofos | 0.9990 | 8% | 94% | 0.9990 | 17% | 76% |
| Endosulfan sulfate | 0.9986 | 6% | 115% | 0.9996 | 13% | 109% |
| Carbophenothion | 0.9963 | 6% | 92% | 0.9990 | 11% | 74% |
| 4,4'-DDT | 0.9987 | 2% | 147% | 0.9990 | 19% | 94% |
| Endrin ketone | 0.9962 | 12% | 95% | 0.9985 | 11% | 83% |
| Methoxychlor | 0.9966 | 15% | 136% | 0.9979 | 18% | 99% |
| Leptophos | 0.9953 | 19% | 59% | 0.9967 | 6% | 57% |

Table 1. Linearity, precision, and recovery of the organochlorine and organophosphate pesticides in tomato juice and wine based on the fully automated QuEChERS sample preparation workflow and GC-MS/MS analysis.

Sample Evaluation

The organochlorine and organophosphate pesticide recoveries were evaluated based on pre- and post-spike of pesticide standards into the tomato juice and red wine samples, respectively. Matrix match calibration curves were generated by using the PAL RTC System to spike seven different concentrations ranging from 1 ng/mL to 100 ng/mL automatically into the cleaned tomato juice and red wine samples (prepared by the automated QuEChERS).

A series of five to six samples, spiked with 10 ng/mL organochlorine and organophosphate pesticide standards, were processed the full automated QuEChERS workflow to determine the recovery and precision based on the matrix match calibration curve. The detailed results of matrix match calibration curve linearity, precision and recovery of this fully automated QuEChERS sample preparation of tomato juice and red wine are listed in Table 1.

Conclusion

A fully automated QuEChERS based on the PAL RTC system is stable and reliable in the analysis of organochlorine and organophosphate pesticides from tomato juice and red wine.

References

[1] QuEChERS Home Page <https://www.quechers.com/index.php> (accessed Sep 7, 2020)

For the full report please request from GLIM@CTC.CH.